



Contents lists available at ScienceDirect

Clinical Nutrition

journal homepage: <http://www.elsevier.com/locate/clnu>

ESPGHAN/ESPEN/ESPR guidelines on pediatric parenteral nutrition: Carbohydrates

D. Mesotten ^{a,*}, K. Joosten ^b, A. van Kempen ^c, S. Verbruggen ^b, the ESPGHAN/ESPEN/ESPR/CSPEN working group on pediatric parenteral nutrition¹

^a University Hospitals Leuven, Department of Intensive Care Medicine, KU Leuven, Leuven, Belgium

^b Sophia Children's Hospital, Department of Pediatrics and Pediatric Surgery, Subdivision Intensive Care, Erasmus MC, Rotterdam, The Netherlands

^c Department of Pediatrics and Neonatology, OLVG, Amsterdam, The Netherlands

ARTICLE INFO

Article history:

Received 29 May 2018

Accepted 29 May 2018

* Corresponding author.

E-mail address: walter.mihatsch@gmx.de (D. Mesotten).

¹ ESPGHAN/ESPEN/ESPR/CSPEN working group on Pediatric Parenteral Nutrition: BRAEGGER Christian, University Children's Hospital, Zurich, Switzerland; BRONSKY Jiri, University Hospital Motol, Prague, Czech Republic; CAI Wei, Shanghai Jiao Tong University, Shanghai, China; CAMPOY Cristina, Department of Paediatrics, School of Medicine, University of Granada, Granada, Spain; CARNIELLI Virgilio, Polytechnic University of Marche, Ancona, Italy; DARMAUN Dominique, Université de Nantes, Nantes, France; DECSI Tamás, Department of Pediatrics, University of Pécs, Pécs, Hungary; DOMELLÖF Magnus, Department of Clinical Sciences, Pediatrics, Umeå University, Sweden; EMBLETON Nicholas, Newcastle University, Newcastle upon Tyne, The United Kingdom; FEWTRELL Mary, UCL Great Ormond Street Institute of Child Health, London, UK; FIDLER MIS Nataša, University Medical Centre Ljubljana, Ljubljana, Slovenia; FRANZ Axel, University Children's Hospital, Tuebingen, Germany; GOULET Olivier, University Sordonne-Paris-Cité; Paris-Descartes Medical School, Paris, France; HARTMAN Corina, Schneider Children's Medical Center of Israel, Petach Tikva, Israel and Carmel Medical Center, Haifa Israel; HILL Susan, Great Ormond Street Hospital for Children, NHS Foundation Trust and UCL Institute of Child Health, London, United Kingdom; HOJSAK Iva, Children's Hospital Zagreb, University of Zagreb School of Medicine, University of J. J. Strossmayer School of Medicine Osijek, Croatia; IACOBELLI Silvia, CHU La Réunion, Saint Pierre, France; JOCHUM Frank, Ev. Waldkrankenhaus Spandau, Berlin, Germany; JOOSTEN, Koen, Department of Pediatrics and Pediatric Surgery, Intensive Care, Erasmus MC-Sophia Children's Hospital, Rotterdam, The Netherlands; KOLACEK Sanja, Children's Hospital, University of Zagreb School of Medicine, Zagreb, Croatia; KOLETZKO Berthold, k LMU – Ludwig-Maximilians-Universität Munich, Dr. von Hauner Children's Hospital, Munich, Germany; KSIAZYK Janusz, Department of Pediatrics, Nutrition and Metabolic Diseases, The Children's Memorial Health Institute, Warsaw; LAPILLONNE Alexandre, Paris-Descartes University, Paris, France; LOHNER Szimonetta, Department of Pediatrics, University of Pécs, Pécs, Hungary; MESOTTEN Dieter, KU Leuven, Leuven, Belgium; MIHÁLYI Krisztina, Department of Pediatrics, University of Pécs, Pécs, Hungary; MIHATSCH Walter A., Ulm University, Ulm, and Helios Hospital, Pforzheim, Germany; MIMOUNI Francis, Department of Pediatrics, Division of Neonatology, The Wilf Children's Hospital, the Shaare Zedek Medical Center, Jerusalem, and the Tel Aviv University, Tel Aviv, Israel; MØLGAARD Christian, Department of Nutrition, Exercise and Sports, University of Copenhagen, and Paediatric Nutrition Unit, Rigshospitalet, Copenhagen, Denmark; MOLTU Sissel J., Oslo University Hospital, Oslo, Norway; NOMAYO Antonia, Ev. Waldkrankenhaus Spandau, Berlin, Germany; PICAUD Jean Charles, Laboratoire CarMEN, Claude Bernard University Lyon 1, Hôpital croix rouge, Lyon, France; PRELL Christine, LMU – Ludwig-Maximilians-Universität Munich, Dr. von Hauner Children's Hospital, Munich, Germany; PUNTIS John, The General Infirmary at Leeds, Leeds, UK; RISKIN Arie, Bnai Zion Medical Center, Rappaport Faculty of Medicine, Technion, Haifa, Israel; SAENZ DE PIPAON Miguel, Department of Neonatology, La Paz University Hospital, Red de Salud Materno Infantil y Desarrollo – SAMID, Universidad Autónoma de Madrid, Madrid, Spain; SENTERRE Thibault, CHU de Liège, CHR de la Citadelle, Université de Liège, Belgium; SHAMIR Raanan, Schneider Children's Medical Center of Israel, Petach Tikva, Israel; Tel Aviv University, Tel Aviv, Israel; SIMCHOWITZ Venetia, Great Ormond Street NHS Trust, London, The United Kingdom; SZITANYI Peter, General University Hospital, First Faculty of Medicine, Charles University in Prague, Czech Republic; TABBERS Merit M., Emma Children's Hospital, Amsterdam UMC, Amsterdam, The Netherlands; VAN DEN AKKER Chris H.B., Emma Children's Hospital, Amsterdam UMC, Amsterdam, The Netherlands; VAN GOUDOEVER Johannes B., Emma Children's Hospital, Amsterdam UMC, Amsterdam, The Netherlands; VAN KEMPEN Anne, OLVG, Amsterdam, the Netherlands; VERBRUGGEN Sascha, Department of Pediatrics and Pediatric Surgery, Intensive Care, Erasmus MC-Sophia Children's Hospital, Rotterdam, The Netherlands; WU Jiang, Xin Hua Hospital, Shanghai, China; YAN Weihui, Department of Gastroenterology and Nutrition, Xinhua Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai, China.

<http://dx.doi.org/10.1016/j.clnu.2018.06.947>

0261-5614/© 2018 European Society for Clinical Nutrition and Metabolism. Published by Elsevier Ltd. All rights reserved.

Please cite this article in press as: Mesotten D, et al., ESPGHAN/ESPEN/ESPR guidelines on pediatric parenteral nutrition: Carbohydrates, Clinical Nutrition (2018), <http://dx.doi.org/10.1016/j.clnu.2018.06.947>

Table: Recommendations for carbohydrates

R 5.1	The amount of glucose to be provided by PN should be guided by [1] the balance between meeting energy needs and the risks of overfeeding/excess glucose load [2], phase of illness (acute, stable, recovery/growing) [3], macronutrient supply by enteral and parenteral nutrition, and [4] glucose administered outside enteral and parenteral nutrition, e.g. with medication (GPP, conditional recommendation)		
R 5.2	Excessive glucose intake should be avoided because it may be responsible for hyperglycemia (LoE 1–, RG A, strong recommendation), causes increased lipogenesis and fat tissue deposition together with subsequent liver steatosis and enhanced production of VLDL triglycerides by the liver (LOE 2+, RG B, strong recommendation), and may cause increased CO ₂ production and minute ventilation (LoE 2+, RG B, strong recommendation)		
R 5.3	Glucose intake does not lower protein catabolism in the acute phase of critical illness (LoE 1–, RG A, strong recommendation)		
R 5.4	Recommended parenteral glucose supply in (pre)term newborns in mg/kg per min (g/kg per day) (LoE 2+, RG B, conditional recommendation)		
		Day 1	Day 2 onwards
		Start with	Increase gradually over 2–3 days to
	Preterm newborn	4–8 (5.8–11.5)	Target 8–10 (11.5–14.4) Min 4 (5.8); max 12 (17.3)
	Term newborn	2.5–5 (3.6–7.2)	Target 5–10 (7.2–14.4) Min 2.5 (3.6); max 12 (17.3)
R 5.5	Newborns < 28 days of age, who have an episode of acute illness such as infection or sepsis, should temporarily receive the carbohydrate supply of day 1 (R5.4), guided by the blood glucose levels (GPP, conditional recommendation)		
R 5.6	Recommended parenteral glucose supply in infants and children according to body weight and phase of illness. The units are mg/kg/min (g/kg per day) (LoE 1+, RG A, strong recommendation)		
		Acute phase	Stable phase
	28 d–10 kg	2–4 (2.9–5.8)	4–6 (5.8–8.6)
	11–30 kg	1.5–2.5 (2.2–3.6)	2–4 (2.8–5.8)
	31–45 kg	1–1.5 (1.4–2.2)	1.5–3 (2.2–4.3)
	>45 kg	0.5–1 (0.7–1.4)	1–2 (1.4–2.9)
		Recovery phase	
		6–10 (8.6–14)	3–6 (4.3–8.6)
		3–4 (4.3–5.8)	2–3 (2.9–4.3)
		Acute phase = resuscitation phase when the patient requires vital organ support (sedation, mechanical ventilation, vasopressors, fluid resuscitation). Stable phase = patient is stable on, or can be weaned, from this vital support. Recovery phase = patient who is mobilizing.	
R 5.7	Blood glucose measurements should preferably be performed on equipment validated for use such as blood gas analysers (LoE 2+, RG B, strong recommendation)		
R 5.8	Hyperglycaemia >8 mmol/L (145 mg/dL) should be avoided in paediatric ICU patients because of increased morbidity and mortality (LoE 1+, RG A, strong recommendation)		
R 5.9	In children in the PICU, repetitive blood glucose levels >10 mmol/L (180 mg/dL) should be treated with continuous insulin infusion (LoE 1+, RG A, strong recommendation)		
R 5.10	Hyperglycaemia >8 mmol/L (145 mg/dL) should be avoided in neonatal ICU patients because it is associated with increased morbidity and mortality (LoE 2–, RG B, strong recommendation)		
R 5.11	In neonates in the NICU, repetitive blood glucose levels >10 mmol/L (180 mg/dL) should be treated with insulin therapy, when reasonable adaptation of glucose infusion rate has been insufficient (LoE 2+, RG 0, conditional recommendation)		
R 5.12	Repetitive and/or prolonged hypoglycaemia ≤2.5 mmol/L (45 mg/dL) should be avoided in all ICU patients (extrapolated LoE 2+, RG 0, strong recommendation)		

1. Methods

Literature Search

Medline search, Pub-Med search, Embase, expert search

Search conducted on 30.11.2014 and on 17.09.2016

Timeframe: publications from <1946 to 17.09.2016>.

Type of publications: original papers, meta-analyses and overviews

Key words: children, parenteral nutrition, glucose, carbohydrate, energy-resource, insulin, critical illness

Language: English

2. Introduction

R 5.1 The amount of glucose to be provided by PN should be guided by [1] the balance between meeting energy needs and the risks of overfeeding/excess glucose load [2], phase of illness (acute, stable, recovery/growing) [3], macronutrient supply by enteral and parenteral nutrition, and [4] glucose administered outside enteral and parenteral nutrition, e.g. with medication (GPP, conditional recommendation, strong consensus)

Carbohydrates are the main source of energy in nutrition and usually provide 40–60% of the energy supply in western diets. The majority of the carbohydrate derived from a normal diet reaches the body's peripheral tissues as glucose. Glucose is utilised by all

cells and serves as metabolic fuel for muscle, liver, heart, kidneys and gut and as the obligate energy source for brain, renal medulla and erythrocytes. Glucose is the main carbohydrate utilized during foetal life; in the last trimester of pregnancy about 5 mg/kg per min (7 g/kg per day) of glucose crosses the placenta. In parenteral nutrition (PN) carbohydrate is provided as dextrose (D-Glucose), in its monohydrate form. Dextrose usually contributes most to the osmolality of the PN-solution.

Recommendations were established by considering [1] the consequences of excessive glucose intake during PN [2], the rate of glucose production and oxidation and [3] the risk of hypoglycaemia. Energy provision during PN includes the use of intravenous fat emulsions (IVFE) (see Lipids chapter) and intravenous amino acid administration (see Amino acids chapter). Therefore, the recommendations for these macronutrients need to be taken into account in order to meet the energy requirements.

When establishing the lower and upper glucose intake recommendations two important factors have to be considered; respectively cerebral glucose utilization and the effect of glucose intake on protein catabolism [1]. A recommendation for higher glucose intake in the neonatal or paediatric ICU would decrease the risk of hypoglycaemia and presumably provide more energy for protein anabolism and growth. However, whole body glucose metabolism in neonates and children is highly modified during (acute) critical illness [2–4]. During acute illness protein catabolism is not modified with increasing glucose intake, while hyperglycaemia, which occurs more frequently during this phase, might be as undesirable as hypoglycaemia [5–7]. Therefore, the basis for glucose intake

recommendation in the acute, critically ill neonate or child deserves a separate approach.

Glucose metabolism is influenced by age, acute illness, nutritional state and the concomitant provision of other macronutrients. Hence, the amount of glucose to be provided by PN should be guided by [1] the balance between meeting energy needs and the risks of overfeeding/excess glucose load [2], phase of illness (acute, stable, recovery/growing) [3], macronutrient supply from enteral and parenteral nutrition, and [4] glucose administered outside enteral and parenteral nutrition, e.g. with medication.

The statements and recommendations that follow should be taken into consideration when treating a (critically) ill child or neonate who cannot be enterally fed during the acute and/or stable phase of his illness. Neonates and children with a (suspected) underlying metabolic disorder require specific carbohydrate intakes, which are not covered in this chapter.

3. Consequences of overfeeding with glucose

-
- R 5.2** Excessive glucose intake should be avoided because it may be responsible for hyperglycemia (LoE 1–, RG A, strong recommendation), causes increased lipogenesis and fat tissue deposition together with subsequent liver steatosis and enhanced production of VLDL triglycerides by the liver (LOE 2+, RG B, strong recommendation), and may cause increased CO₂ production and minute ventilation (LoE 2+, RG B, strong recommendation, strong consensus)
- R 5.3** Glucose intake does not lower protein catabolism in the acute phase of critical illness (LoE 1–, RG A, strong recommendation, strong consensus)
-

When glucose is administered in excess of the amount that can be directly oxidized for energy production and glycogen synthesis, the excess is directed to lipogenesis, thus promoting fat deposition [8,9]. Restoration or accumulation of fat stores may be a nutritional goal in infants and children with (severe) malnutrition or rapid growth, by providing more lipids rather than by excessive carbohydrate administration. Excessive fat deposition and dyslipidaemia may be deleterious, especially during the acute phase of critical illness [10]. The conversion of glucose into lipids partially accounts for the increase in energy expenditure observed with high rates of glucose infusion [11]. Excessive glucose intake as well as total energy delivery and amino acid intake, increases CO₂ production and minute ventilation [12–14]. Excessive glucose intake may also impair liver function especially by inducing steatosis, while its contribution to the development of cholestasis is not clearly established [15,16]. Studies in healthy adults suggest that high carbohydrate feeding leads to an increase in total very-low-density lipoprotein (VLDL) triglyceride secretion rate from de novo synthesis, primarily due to stimulation of the secretion of preformed fatty acids (FA) [17]. These results imply that the liver derives its energy from carbohydrate oxidation rather than from FA oxidation. FA taken up by the liver are channelled into VLDL triglycerides. Hepatic steatosis results when export of the VLDL triglycerides does not keep pace with triglyceride production [17,18]. High carbohydrate intake, both in hypercaloric as well as eucaloric conditions, leads to lipogenesis [19,20].

Furthermore, high carbohydrate intake induces insulin resistance through activation of the transcription factor ChREBP (carbohydrate response element binding protein) to protect the liver from glucose overload, which will lead to a counterproductive increase in hepatic glucose production [21]. Critical illness causes dyslipidaemia, characterized by increased triglycerides and VLDL, and hypocholesterolaemia [10,22,23]. Although these pathways

have not been thoroughly studied in critically ill neonates or children, dyslipidaemia has been observed in septic children [24]. Therefore, excess glucose intake may exacerbate critical illness related dyslipidaemia in children as in adults.

Another concern of parenteral glucose overfeeding is its association with hyperglycaemia. In critically ill children this is caused by insulin resistance as well as beta-cell dysfunction [25,26]. The consequences and management of hyperglycaemia in critically ill children are discussed in the final paragraph of this chapter.

Adding lipid emulsions and amino acid infusions allow the energy input to be diversified, and glucose intake to be decreased, while maintaining adequate energy intake [27]. In preterm newborns, protein metabolism is influenced by the amount and composition of energy intake [28,29].

The glucose intake recommendations in the former guidelines did not cater for acute critical illness [30]. Under these circumstances, the administration of total caloric and glucose amounts appropriate for healthy, growing infants and children may induce hyperglycaemia and other metabolic derangements [5,31]. Decreased energy recommendations in the acute phase of critical illness (chapter 1) allow the parenteral glucose intake to be lowered. The amount of glucose and/or energy intake does not impact protein metabolism in the acute post-operative phase [6,7]. Reduced glucose intake in these critically ill infants safely lowered high blood glucose levels, despite an increased endogenous glucose production [31,32]. A study in burned children (age 7.3 ± 5.4 y) also showed that judicious use of parenteral nutrition within one week of injury by capping glucose intake at 5–7 mg/kg/min was safe and effective, while minimizing complications of PN [33]. When a patient is recovering, insulin resistance will decrease and glucose metabolism will improve. This will allow a higher glucose supply, necessary for rehabilitation and growth.

4. Rate of endogenous glucose production and rate of glucose oxidation

The efficiency with which glucose is utilised should guide the upper limit of carbohydrate supply, while the lower limit is defined by the risk of hypoglycaemia. The majority of quantitative estimates of production and oxidation of glucose have been performed using stable isotopic tracers and indirect calorimetry (IC) in healthy term or preterm newborns. Stable isotope studies cannot be used at the bedside and IC has several limitations. Furthermore, IC uses a Respiratory Quotient >1 as marker of excessive glucose intake, but this has not been validated. Rate of glucose oxidation (RGO) and endogenous rate of glucose production (RGP) can be measured with stable isotopes. Exogenous glucose delivered in excess of the rate of glucose oxidation (RGO) may enter non-oxidative pathways and is unlikely to improve energy balance. Decreasing or stopping endogenous glucose production would be a normal physiological response. When exogenous glucose is insufficient this would increase the RGP, however this could be insufficient to prevent hypoglycaemia. Again, these responses are affected by age as well by the phase of illness.

4.1. Endogenous glucose production in preterm infants

In preterm infants RGP, gluconeogenesis and glycogenolysis have been studied under different nutritional circumstances, showing that RGP in preterm infants is influenced by IV glucose and PN. RGP increased in preterm infants when the exclusive IV exogenous glucose administration was diminished from 6 to 4 mg/kg per min. Nevertheless, the increased RGP was not enough to

prevent a drop in plasma glucose concentration [39]. Gluconeogenesis is responsible for about 31% of RGP in fasting, healthy full term newborns [40] and for up to 75% in healthy preterms receiving IV glucose or PN [39,41]. RGP and gluconeogenesis can be stimulated in preterm infants by administration of glycerol, IV lipids or PN [39,42–44], but not by the administration of alanine [45]. Glucagon increases glucose production from glycogenolysis in preterm infants. Nevertheless, the response is low, especially considering their increased needs [46]. These studies show that preterm infants are capable of glucose production and gluconeogenesis. However, production capacity is limited and therefore they depend both on exogenous glucose and PN components to maintain glucose homeostasis and avoid hypoglycaemia.

On the other hand, several studies showed that in extremely preterm neonates (24–29 weeks) endogenous glucose production and gluconeogenesis on day 3–4 were not affected by the glucose infusion rate or blood glucose levels [41,42,47]. In contrast, in moderately preterm neonates (31 ± 1.5 weeks) the endogenous glucose production on day 8 was suppressed completely by parenteral glucose intake [48]. These studies suggest that the inability to suppress glucose production or gluconeogenesis may contribute to the risk of hyperglycaemia in extremely preterm infants.

4.2. Endogenous glucose production in older infants and children

The basal rate of endogenous glucose production (RGP) varies from 2 mg/kg per min (2.9 g/kg per day) in adults, to 8 mg/kg per min (11.5 g/kg per day) in preterm infants [39,49]. The RGP is maximal during the postnatal period and decreases gradually with age [46]. Few studies are available for infants and children, and even fewer during acute critical illness. In post-surgical critically ill infants (5–10 months of age) reducing parenteral glucose intake in the acute phase to 2.5 mg/kg per min lowered high glycaemic levels and increased the RGP, primarily through increased glycogenolysis [31,32].

4.3. Rate of glucose oxidation

During PN, the rate of parenteral glucose delivery should not exceed the maximum rate of glucose oxidation (RGO). Only three studies have measured RGO in children, showing significant differences among patients according to their age and clinical status. In appropriate for gestational age preterm infants, the RGO is 6–8 mg/kg per min (8.6–11.5 g/kg per day) [50,51]. In term infants after surgery or infants on long-term PN, the maximal RGO is about 12 mg/kg per min (17.2 g/kg per day) [52,53]. In contrast, a small study in critically burned children (1–11 y) demonstrated the maximal RGO (3.8 mg/kg per min or 5.5 g/kg per day) to be at a glucose intake of 5 mg/kg per min [54].

4.4. General recommendations for parenteral carbohydrate intake

R 5.4 Recommended parenteral glucose supply in (pre)term newborns in mg/kg per min (g/kg per day) (LoE 2+, RG B, conditional recommendation, strong consensus)

	Day 1	Day 2 onwards
	Start with	Increase gradually over 2–3 days to
Preterm newborn	4–8 (5.8–11.5)	Target 8–10 (11.5–14.4) Min 4 (5.8); max 12 (17.3)
Term newborn	2.5–5 (3.6–7.2)	Target 5–10 (7.2–14.4) Min 2.5 (3.6); max 12 (17.3)

R 5.5 Newborns < 28 days of age, who have an episode of acute illness such as infection or sepsis, should temporarily receive the carbohydrate supply of day 1 (R5.4), guided by the blood glucose levels (GPP, conditional recommendation, strong consensus)

R 5.6 Recommended parenteral glucose supply in infants and children according to body weight and phase of illness. The units are mg/kg/min (g/kg per day) (LoE 1+, RG A, strong recommendation, strong consensus)

	Acute phase	Stable phase	Recovery phase
28 d–10 kg	2–4 (2.9–5.8)	4–6 (5.8–8.6)	6–10 (8.6–14)
11–30 kg	1.5–2.5 (3.6–2.9)	2–4 (2.8–5.8)	3–6 (4.3–8.6)
31–45 kg	1–1.5 (1.4–2.2)	1.5–3 (2.2–4.3)	3–4 (4.3–5.8)
> 45 kg	0.5–1 (0.7–1.4)	1–2 (1.4–2.9)	2–3 (2.9–4.3)

Acute phase = resuscitation phase when the patient requires vital organ support (sedation, mechanical ventilation, vasopressors, fluid resuscitation).

Stable phase = patient is stable on, or can be weaned, from this vital support.

Recovery phase = patient who is mobilizing.

The phase of critical illness plays a role in the energy requirement (also see chapter Energy) and hence also in the carbohydrate supply [55]. A recent large international multicentre randomised controlled trial in 1440 critically ill children, including term neonates, (PEPaNIC study) compared whether a strategy of withholding parenteral nutrition up to day 8 in the PICU (late parenteral nutrition) was clinically superior to early initiation of supplemental PN (initiated within 24 h after admission) [37, 38]. It was shown that withholding parenteral nutrition for 1 week while administering micronutrients intravenously was clinically superior to providing early parenteral nutrition to supplement insufficient enteral nutrition. No parenteral nutrition for 1 week significantly reduced the number of new infections, the time on a ventilator, kidney failure and increased the likelihood of earlier live discharge from the PICU and the hospital with decreased direct medical costs [34–36,40]. Based on the above statements we propose that most likely lower amounts of energy/carbohydrate should be given to acutely critically ill children. This acute phase of critical illness (first hours to days) only covers the resuscitation phase when the unstable patient requires vital organ support (sedation, mechanical ventilation, vasopressors, fluid resuscitation). When a patient has been stabilised on, or can be weaned from, this vital support, he/she is in the stable phase. When the child is mobilising, it is called the recovery phase [55]. In the recovery phase more energy/carbohydrates should be provided, which should be further increased in the recovery phase in order to achieve growth.

In (preterm) newborns energy/carbohydrate amounts are gradually increased over the first postnatal days. Carbohydrate intake is determined by energy requirements, blood glucose levels and – after the nadir in postnatal weight loss – growth. The blood glucose level is an important determinant for glucose supply on the first postnatal day. Thereafter the glucose intake is increased stepwise over the next 2–3 days, usually up to 10 mg/kg per min (14.4 g/kg per day) in order to allow growth. Parenteral carbohydrate intake should preferably not exceed 12 mg/kg per min (17.3 g/kg per day) and generally not be lower than 4 mg/kg per min (5.8 g/kg per day) in preterm infants or 2.5 mg/kg per min (3.6 g/kg per day) in term newborns.

Carbohydrate intake must be individualized, especially in newborn infants with specific problems, e.g. hypo- or hyperglycaemia, severe perinatal asphyxia (as concomitant hypoglycaemia may exacerbate brain damage), hyperinsulinaemia, and newborns on (long-term) PN with lipid intolerance or insufficient growth. Finally, as stated before, these statements and recommendations are not applicable to neonates and children with a (suspected) metabolic disorder.

5. Dysglycemia and blood glucose management

5.1. Blood glucose measurements

R 5.7 **Blood glucose measurements should preferably be performed on blood gas analysers (LoE 2+, RG B, strong recommendation, strong consensus)**

Blood glucose management starts with measuring blood glucose levels. These measurements should be accurate and accessible for bedside nurses and doctors at the bedside. Due to the use of capillary blood, anaemia and drugs that interfere with the enzymatic reaction of the blood glucose measurement such as ascorbic acid and acetaminophen, the accuracy of handheld blood glucose meters is less accurate in critically ill patients [56]. In critically ill patients blood glucose levels can be measured most accurately yet still practically on arterial blood using blood gas analysers [57–59]. In patients who do not need an arterial line, handheld blood glucose meters may be used [58,60].

In newborn infants the accuracy of handheld blood glucose meters is still of great concern [60–62]. Factors that influence glucose measurements are (amongst others) high haemoglobin levels and high bilirubin levels [62–64]. Despite this, handheld blood glucose meters are frequently used in daily clinical practice since they provide very rapid results. Standard laboratory testing is not preferable because of the delay in obtaining a result and the possibility of falsely low results due to ongoing glycolysis in the sample, if appropriate pre-analytical guidelines are neglected [65]. At present, the best method combining quick results and accuracy is delivered by blood gas analysers with glucose modules for blood glucose measurements in newborn infants [66,67].

5.2. Hyperglycaemia

R 5.8 **Hyperglycaemia >8 mmol/L (145 mg/dL) should be avoided in paediatric ICU patients because of increased morbidity and mortality (LoE 1+, RG A, strong recommendation, strong consensus)**

R 5.9 **In PICU, repetitive blood glucose levels >10 mmol/L (180 mg/dL) should be treated with continuous insulin infusion (LoE 1+, RG A, strong recommendation, strong consensus)**

R 5.10 **Hyperglycaemia >8 mmol/L (145 mg/dL) should be avoided in neonatal ICU patients because it is associated with increased morbidity and mortality (LoE 2–, RG B, strong recommendation, strong consensus)**

R 5.11 **In NICU, repetitive blood glucose levels >10 mmol/L (180 mg/dL) should be treated with insulin therapy, when reasonable adaptation of glucose infusion rate has been insufficient (LoE 2+, RG 0, conditional recommendation, strong consensus)**

In preterm infants, the most common definition of hyperglycaemia is a blood glucose level exceeding 10 mmol/L (180 mg/dL) [68] and this has been associated with increased morbidity [69–73]. Insulin therapy in (preterm) newborns is effective in treating or preventing hyperglycaemia, but also leads to an increased incidence of hypoglycaemia. There is no evidence for recommending tight blood glucose management in the NICU [74]. Hence, insulin therapy at a low starting dose is preferred and only when reasonable adaptation of the glucose infusion rate is insufficient to control neonatal hyperglycaemia [75,76].

In critically ill children, hyperglycaemia has consistently been associated with increased morbidity and mortality [77–81]. Malnourished children with hyperglycaemia have a greater risk of

mortality than well-nourished patients [82]. The definitions for hyperglycaemia range from blood glucose levels above 7 mmol/L (126 mg/dL) [83] to levels above 8.3 mmol/L (150 mg/dL) [84]. In a single-centre RCT, tight blood glucose management, to levels between 2.8 and 4.4 mmol/L (50–80 mg/dL) in infants and between 3.9 and 5.6 mmol/L (70–100 mg/dL) in children, reduced the incidence of nosocomial infections, shortened length of stay in the ICU and lowered mortality rate [85]. However, a quarter of the children in the intervention group experienced at least one episode of hypoglycaemia below 2.2 mmol/L (40 mg/dL). Also in severely burned paediatric patients, intensive insulin therapy decreased morbidity [86]. Blood glucose control to a slightly higher target range than the study by Vlasselaers et al. did not result in a better outcome in multicentre trials, in comparison with the control group in which insulin treatment was only started in case of excessive hyperglycaemia [87,88]. A meta-analysis of these four trials revealed that tight blood glucose control in critically ill children does not decrease mortality, but reduces new infections. Yet, tight blood glucose control is strongly associated with a higher incidence of hypoglycaemia [89].

5.3. Hypoglycaemia

R 5.12 **Repetitive and/or prolonged hypoglycaemia ≤ 2.5 mmol/L (45 mg/dL) should be avoided in all ICU patients (extrapolated LoE 2+, RG 0, strong recommendation, strong consensus)**

In critically ill children hypoglycaemia is defined as a blood glucose level below 2.8 mmol/L (50 mg/dL) [90] or below 3.3 mmol/L (60 mg/dL) [91]. A recent systematic review and meta-analysis proposed to define hypoglycaemia as 2.2–2.5 mmol/L (<40–45 mg/dL) in newborns and 3.3–3.6 mmol/L (<60–65 mg/dL) in children [90]. The association between hypoglycaemia and mortality risk is less robust in critically ill children, since severity of illness and age may be important confounders [90,92]. Also the long term consequences of a brief period of low glucose levels, that are not associated with clinical signs, remain uncertain. Four years after study inclusion in the trial on tight blood glucose management and being exposed to hypoglycaemia, the children who underwent tight blood glucose control did not show impaired neurocognitive development [92]. Studies on the effect of hypoglycaemia in the postnatal period on subsequent neurodevelopment are mostly of poor methodological quality and so far could not provide a valid estimate [93]. In preterm newborns a large cohort study reported impaired motor and cognitive development at 18 months [94], but found no differences in developmental progress or physical disability 15 years after recurrent low blood glucose levels (≤ 2.5 mmol/L) in the first 10 days after birth [95]. In a more recent cohort study neonatal (≥ 35 weeks) hypoglycaemia was not associated with impaired neurological outcome at two years when treated to maintain blood glucose concentrations of at least 2.6 mmol/L (47 mg/dL) [96]. In (preterm) newborns the suggested blood glucose operational threshold concentrations at which clinicians should consider intervention are: a single measurement of blood glucose <1 mmol/L (18 mg/dL); blood glucose level <2 mmol/L (36 mg/dL) which remains below the same value at the next measurement; or a single measurement of <2.5 mmol/L (45 mg/dL) in a newborn with abnormal clinical signs [97]. Certainly newborns with risk factors for hypoglycaemia, such as premature birth, low birth weight and perinatal asphyxia, require close monitoring and management of their blood glucose levels [98].

Conflict of interest

None declared.

References

- [1] Kalhan SC, Kilic I. Carbohydrate as nutrient in the infant and child: range of acceptable intake. *Eur J Clin Nutr* 1999;53(Suppl. 1):S94–100.
- [2] Lang CH, Frost RA, Vary TC. Regulation of muscle protein synthesis during sepsis and inflammation. *Am J Physiol Endocrinol Metab* 2007;293(2):E453–9.
- [3] Magnoni S, Tedesco C, Carbonara M, Pluderi M, Colombo A, Stocchetti N. Relationship between systemic glucose and cerebral glucose is preserved in patients with severe traumatic brain injury, but glucose delivery to the brain may become limited when oxidative metabolism is impaired: implications for glycemic control. *Crit Care Med* 2012;40(6):1785–91.
- [4] Vespa P, McArthur DL, Stein N, Huang SC, Shao W, Filippou M, et al. Tight glycemic control increases metabolic distress in traumatic brain injury: a randomized controlled within-subjects trial. *Crit Care Med* 2012;40(6):1923–9.
- [5] Suresh D, Athanassaki I, Jeha GS, Heptulla RA. Total parenteral nutrition associated with severe insulin resistance following hematopoietic stem cell transplantation in patients with hemophagocytic syndrome: report on two cases. *Pediatr Diabetes* 2010;11(1):70–3.
- [6] Verbruggen SC, Schierbeek H, Coss-Bu J, Joosten KF, Castillo L, van Goudoever JB. Albumin synthesis rates in post-surgical infants and septic adolescents; influence of amino acids, energy, and insulin. *Clin Nutr* 2011;30(4):469–77.
- [7] Geukers VG, Li Z, Ackermans MT, Bos AP, Jinfeng L, Sauerwein HP. High-carbohydrate/low-protein-induced hyperinsulinemia does not improve protein balance in children after cardiac surgery. *Nutrition* 2012;28(6):644–50.
- [8] Robin AP, Carpentier YA, Askanazi J, Nordenstrom J, Kinney JM. Metabolic consequences of hypercaloric glucose infusions. *Acta Chir Belg* 1981;80(2–3):133–40.
- [9] Koretz RL, Lipman TO, Klein S, American Gastroenterological A. AGA technical review on parenteral nutrition. *Gastroenterology* 2001;121(4):970–1001.
- [10] Mesotten D, Swinnen JV, Vanderhoydonc F, Wouters PJ, Van den Berghe G. Contribution of circulating lipids to the improved outcome of critical illness by glycemic control with intensive insulin therapy. *J Clin Endocrinol Metab* 2004;89(1):219–26.
- [11] Elwyn DH, Askanazi J, Kinney JM, Gump FE. Kinetics of energy substrates. *Acta Chir Scand Suppl* 1981;507:209–19.
- [12] Talpers SS, Romberger DJ, Bunce SB, Pingleton SK. Nutritionally associated increased carbon dioxide production. Excess total calories vs high proportion of carbohydrate calories. *Chest* 1992;102(2):551–5.
- [13] Askanazi J, Weissman C, LaSala PA, Milic-Emili J, Kinney JM. Effect of protein intake on ventilatory drive. *Anesthesiology* 1984;60(2):106–10.
- [14] Rodriguez JL, Askanazi J, Weissman C, Hensle TW, Rosenbaum SH, Kinney JM. Ventilatory and metabolic effects of glucose infusions. *Chest* 1985;88(4):512–8.
- [15] Burke JF, Wolfe RR, Mullany CJ, Mathews DE, Bier DM. Glucose requirements following burn injury. Parameters of optimal glucose infusion and possible hepatic and respiratory abnormalities following excessive glucose intake. *Ann Surg* 1979;190(3):274–85.
- [16] Tulikoura I, Huikuri K. Morphological fatty changes and function of the liver, serum free fatty acids, and triglycerides during parenteral nutrition. *Scand J Gastroenterol* 1982;17(2):177–85.
- [17] Aarsland A, Chinkes D, Wolfe RR. Contributions of de novo synthesis of fatty acids to total VLDL-triglyceride secretion during prolonged hyperglycemia/hyperinsulinemia in normal man. *J Clin Invest* 1996;98(9):2008–17.
- [18] Klein CJ, Stanek GS, Wiles 3rd CE. Overfeeding macronutrients to critically ill adults: metabolic complications. *J Am Diet Assoc* 1998;98(7):795–806.
- [19] Schwarz JM, Neese RA, Turner S, Dare D, Hellerstein MK. Short-term alterations in carbohydrate energy intake in humans. Striking effects on hepatic glucose production, de novo lipogenesis, lipolysis, and whole-body fuel selection. *J Clin Invest* 1995;96(6):2735–43.
- [20] Hudgins LC, Hellerstein M, Seidman C, Neese R, Diakun J, Hirsch J. Human fatty acid synthesis is stimulated by a eucaloric low fat, high carbohydrate diet. *J Clin Invest* 1996;97(9):2081–91.
- [21] Agius L. High-carbohydrate diets induce hepatic insulin resistance to protect the liver from substrate overload. *Biochem Pharmacol* 2013;85(3):306–12.
- [22] Khovidhunkit W, Kim MS, Memon RA, Shigenaga JK, Moser AH, Feingold KR, et al. Effects of infection and inflammation on lipid and lipoprotein metabolism: mechanisms and consequences to the host. *J Lipid Res* 2004;45(7):1169–96.
- [23] Chien JY, Jerng JS, Yu CJ, Yang PC. Low serum level of high-density lipoprotein cholesterol is a poor prognostic factor for severe sepsis. *Crit Care Med* 2005;33(8):1688–93.
- [24] Vermont CL, den Brinker M, Kakeci N, de Kleijn ED, de Rijke YB, Joosten KF, et al. Serum lipids and disease severity in children with severe meningococcal sepsis. *Crit Care Med* 2005;33(7):1610–5.
- [25] Preissig CM, Rigby MR. Hyperglycaemia results from beta-cell dysfunction in critically ill children with respiratory and cardiovascular failure: a prospective observational study. *Crit Care* 2009;13(1):R27.
- [26] Verbruggen SC, Coss-Bu J, Wu M, Schierbeek H, Joosten KF, Dhar A, et al. Current recommended parenteral protein intakes do not support protein synthesis in critically ill septic, insulin-resistant adolescents with tight glucose control. *Crit Care Med* 2011;39(11):2518–25.
- [27] Vlaardingerbroek H, Vermeulen MJ, Rook D, van den Akker CH, Dorst K, Wattimena JL, et al. Safety and efficacy of early parenteral lipid and high-dose amino acid administration to very low birth weight infants. *J Pediatr* 2013;163(3):638–44 e1–5.
- [28] Macfie J, Smith RC, Hill GL. Glucose or fat as a nonprotein energy source? A controlled clinical trial in gastroenterological patients requiring intravenous nutrition. *Gastroenterology* 1981;80(1):103–7.
- [29] Pineault M, Chessex P, Bisailon S, Brisson G. Total parenteral nutrition in the newborn: impact of the quality of infused energy on nitrogen metabolism. *Am J Clin Nutr* 1988;47(2):298–304.
- [30] Koletzko B, Goulet O, Hunt J, Krohn K, Shamir R. Parenteral Nutrition Guidelines Working G, et al. Guidelines on Paediatric Parenteral Nutrition of the European Society of Paediatric Gastroenterology, Hepatology and Nutrition (ESPGHAN) and the European Society for Clinical Nutrition and Metabolism (ESPEN), supported by the European Society of Paediatric Research (ESPR). *J Pediatr Gastroenterol Nutr* 2005;41(Suppl. 2):S1–87.
- [31] de Betue CT, Verbruggen SC, Schierbeek H, Chacko SK, Bogers AJ, van Goudoever JB, et al. Does a reduced glucose intake prevent hyperglycemia in children early after cardiac surgery? A randomized controlled crossover study. *Crit Care* 2012;16(5):R176.
- [32] Verbruggen SC, de Betue CT, Schierbeek H, Chacko S, van Adrichem LN, Verhoeven J, et al. Reducing glucose infusion safely prevents hyperglycemia in post-surgical children. *Clin Nutr* 2011;30(6):786–92.
- [33] Dylewski ML, Baker M, Prelack K, Weber JM, Hursey D, Lydon M, et al. The safety and efficacy of parenteral nutrition among pediatric patients with burn injuries. *Pediatr Crit Care Med* 2013;14(3):e120–5.
- [34] Vanhorebeek I, Verbruggen S, Casaer MP, Gunst J, Wouters PJ, Hanot J, et al. Effect of early supplemental parenteral nutrition in the paediatric ICU: a preplanned observational study of post-randomisation treatments in the PEPaNIC trial. *Lancet Respir Med* 2017 Jun;5(6):475–83. [http://dx.doi.org/10.1016/S2213-2600\(17\)30186-8](http://dx.doi.org/10.1016/S2213-2600(17)30186-8). Epub 2017 May 15.
- [35] Van Puffelen E, Vanhorebeek I, Joosten KF, Wouters PJ, Van den Berghe G, Verbruggen SC. Early versus late parenteral nutrition in critically ill, term neonates: a preplanned secondary subgroup analysis of the PEPaNIC multicentre, randomised controlled trial. *The Lancet Child & Adolescent Health* 2(7):505–515.
- [36] Van Puffelen E, Polinder S. Cost-effectiveness study of early versus late parenteral nutrition in critically ill children (PEPaNIC): preplanned secondary analysis of a multicentre randomised controlled trial. *Crit Care* 2018 Jan 15;22(1):4. <http://dx.doi.org/10.1186/s13054-017-1936-2>.
- [37] Fivez T, Kerklaan D, Verbruggen S, Vanhorebeek I, Verstraete S, Tibboel D, et al. Impact of withholding early parenteral nutrition completing enteral nutrition in pediatric critically ill patients (PEPaNIC trial): study protocol for a randomized controlled trial. *Trials* 2015;16:202.
- [38] Fivez T, Kerklaan D, Mesotten D, Verbruggen S, Wouters PJ, Vanhorebeek I, et al. Early versus late parenteral nutrition in critically ill children. *N Engl J Med* 2016;374(12):1111–22.
- [39] Van Kempen AA, Romijn JA, Ruiters AF, Ackermans MT, Ender E, Hoekstra JH, et al. Adaptation of glucose production and gluconeogenesis to diminishing glucose infusion in preterm infants at varying gestational ages. *Pediatr Res* 2003;53(4):628–34.
- [40] Kalhan SC, Parimi P, Van Beek R, Gilfillan C, Saker F, Gruca L, et al. Estimation of gluconeogenesis in newborn infants. *Am J Physiol Endocrinol Metab* 2001;281(5):E991–7.
- [41] Chacko SK, Sunehag AL. Gluconeogenesis continues in premature infants receiving total parenteral nutrition. *Arch Dis Child Fetal Neonatal Ed* 2010;95(6):F413–8.
- [42] Chacko SK, Ordonez J, Sauer PJ, Sunehag AL. Gluconeogenesis is not regulated by either glucose or insulin in extremely low birth weight infants receiving total parenteral nutrition. *J Pediatr* 2011;158(6):891–6.
- [43] Sunehag AL, Haymond MW, Schanler RJ, Reeds PJ, Bier DM. Gluconeogenesis in very low birth weight infants receiving total parenteral nutrition. *Diabetes* 1999;48(4):791–800.
- [44] van Kempen AA, van der Crabben SN, Ackermans MT, Ender E, Kok JH, Sauerwein HP. Stimulation of gluconeogenesis by intravenous lipids in preterm infants: response depends on fatty acid profile. *Am J Physiol Endocrinol Metab* 2006;290(4):E723–30.
- [45] van Kempen AA, Romijn JA, Ruiters AF, Ender E, Weverling GJ, Kok JH, et al. Alanine administration does not stimulate gluconeogenesis in preterm infants. *Metab Clin Exp* 2003;52(8):945–9.
- [46] van Kempen AA, Ackermans MT, Ender E, Kok JH, Sauerwein HP. Glucose production in response to glucagon is comparable in preterm AGA and SGA infants. *Clin Nutr* 2005;24(5):727–36.
- [47] Cowett RM, Oh W, Schwartz R. Persistent glucose production during glucose infusion in the neonate. *J Clin Invest* 1983;71(3):467–75.
- [48] Lafeber HN, Sulkers EJ, Chapman TE, Sauer PJ. Glucose production and oxidation in preterm infants during total parenteral nutrition. *Pediatr Res* 1990;28(2):153–7.
- [49] Mitanchez D. Glucose regulation in preterm newborn infants. *Horm Res* 2007;68(6):265–71.

- [50] Forsyth JS, Crichton A. Low birthweight infants and total parenteral nutrition immediately after birth. I. Energy expenditure and respiratory quotient of ventilated and non-ventilated infants. *Arch Dis Child Fetal Neonatal Ed* 1995;73(1):F4–7.
- [51] Sauer PJ, Van Aerde JE, Pencharz PB, Smith JM, Swyer PR. Glucose oxidation rates in newborn infants measured with indirect calorimetry and [^{13}C] glucose. *Clin Sci* 1986;70(6):587–93.
- [52] Jones MO, Pierro A, Hammond P, Nunn A, Lloyd DA. Glucose utilization in the surgical newborn infant receiving total parenteral nutrition. *J Pediatr Surg* 1993;28(9):1121–5.
- [53] Nose O, Tipton JR, Ament ME, Yabuuchi H. Effect of the energy source on changes in energy expenditure, respiratory quotient, and nitrogen balance during total parenteral nutrition in children. *Pediatr Res* 1987;21(6):538–41.
- [54] Sheridan RL, Yu YM, Prelack K, Young VR, Burke JF, Tompkins RG. Maximal parenteral glucose oxidation in hypermetabolic young children: a stable isotope study. *J Parenter Enteral Nutr* 1998;22(4):212–6.
- [55] Van den Berghe G, de Zegher F, Bouillon R. Clinical review 95: acute and prolonged critical illness as different neuroendocrine paradigms. *J Clin Endocrinol Metab* 1998;83(6):1827–34.
- [56] Scott MG, Bruns DE, Boyd JC, Sacks DB. Tight glucose control in the intensive care unit: are glucose meters up to the task? *Clin Chem* 2009;55(1):18–20.
- [57] Kanji S, Buffie J, Hutton B, Bunting PS, Singh A, McDonald K, et al. Reliability of point-of-care testing for glucose measurement in critically ill adults. *Crit Care Med* 2005;33(12):2778–85.
- [58] Finfer S, Wernerman J, Preiser JC, Cass T, Desai T, Hovorka R, et al. Clinical review: consensus recommendations on measurement of blood glucose and reporting glycaemic control in critically ill adults. *Crit Care* 2013;17(3):229.
- [59] Inoue S, Egi M, Kotani J, Morita K. Accuracy of blood-glucose measurements using glucose meters and arterial blood gas analyzers in critically ill adult patients: systematic review. *Crit Care* 2013;17(2):R48.
- [60] Beardsall K. Measurement of glucose levels in the newborn. *Early Hum Dev* 2010;86(5):263–7.
- [61] Woo HC, Tolosa L, El-Metwally D, Viscardi RM. Glucose monitoring in neonates: need for accurate and non-invasive methods. *Arch Dis Child Fetal Neonatal Ed* 2014;99(2):F153–7.
- [62] Tang Z, Lee JH, Louie RF, Kost GJ. Effects of different hematocrit levels on glucose measurements with handheld meters for point-of-care testing. *Arch Pathol Lab Med* 2000;124(8):1135–40.
- [63] Kaplan M, Blondheim O, Alon I, Eylath U, Trestian S, Eidelman AI. Screening for hypoglycemia with plasma in neonatal blood of high hematocrit value. *Crit Care Med* 1989;17(3):279–82.
- [64] Jain R, Myers TF, Kahn SE, Zeller WP. How accurate is glucose analysis in the presence of multiple interfering substances in the neonate? (glucose analysis and interfering substances). *J Clin Lab Anal* 1996;10(1):13–6.
- [65] Hagvik J. Comment on: Bellini C, Serra G, Rizzo D, Mazzella M, Bonioli E. Reliability assessment of glucose measurement by HemoCue analyser in a neonatal intensive care unit. *Clin Chem Lab Med* 2007; 45(11):1549–54. *Clin Chem Lab Med* 2008;46(5):729–30.
- [66] Newman JD, Pecache NS, Barfield CP, Balazs ND. Point-of-care testing of blood glucose in the neonatal unit using the AVL Omni 9 analyser. *Ann Clin Biochem* 2002;39(Pt 5):509–12.
- [67] Peet AC, Kennedy DM, Hocking MD, Ewer AK. Near-patient testing of blood glucose using the Bayer Rapidlab 860 analyser in a regional neonatal unit. *Ann Clin Biochem* 2002;39(Pt 5):502–8.
- [68] Alsweiler JM, Kuschel CA, Bloomfield FH. Survey of the management of neonatal hyperglycaemia in Australasia. *J Paediatr Child Health* 2007;43(9):632–5.
- [69] Ramel SE, Long JD, Gray H, Durrwachter-Erno K, Demerath EW, Rao R. Neonatal hyperglycemia and diminished long-term growth in very low birth weight preterm infants. *J Perinatol* 2013;33(11):882–6.
- [70] Auerbach A, Eventov-Friedman S, Arad I, Peleg O, Bdoлах-Abram T, Bar-Oz B, et al. Long duration of hyperglycemia in the first 96 hours of life is associated with severe intraventricular hemorrhage in preterm infants. *J Pediatr* 2013;163(2):388–93.
- [71] van der Lugt NM, Smits-Wintjens VE, van Zwieten PH, Walther FJ. Short and long term outcome of neonatal hyperglycemia in very preterm infants: a retrospective follow-up study. *BMC Pediatr* 2010;10:52.
- [72] Beardsall K, Vanhaesebrouck S, Ogilvy-Stuart AL, Vanhole C, Palmer CR, Ong K, et al. Prevalence and determinants of hyperglycemia in very low birth weight infants: cohort analyses of the NIRTURE study. *J Pediatr* 2010;157(5): 715–9 e1–3.
- [73] Alaadeen DI, Walsh MC, Chwals WJ. Total parenteral nutrition-associated hyperglycemia correlates with prolonged mechanical ventilation and hospital stay in septic infants. *J Pediatr Surg* 2006;41(1):239–44.
- [74] Ogilvy-Stuart AL, Beardsall K. Management of hyperglycaemia in the preterm infant. *Arch Dis Child Fetal Neonatal Ed* 2010;95(2):F126–31.
- [75] Beardsall K, Vanhaesebrouck S, Ogilvy-Stuart AL, Vanhole C, Palmer CR, van Weissenbruch M, et al. Early insulin therapy in very-low-birth-weight infants. *N Engl J Med* 2008;359(18):1873–84.
- [76] Alsweiler JM, Harding JE, Bloomfield FH. Tight glycaemic control with insulin in hyperglycaemic preterm babies: a randomized controlled trial. *Pediatrics* 2012;129(4):639–47.
- [77] Cochran A, Scaife ER, Hansen KW, Downey EC. Hyperglycemia and outcomes from pediatric traumatic brain injury. *J Trauma* 2003;55(6):1035–8.
- [78] Wintergerst KA, Buckingham B, Gandrud L, Wong BJ, Kache S, Wilson DM. Association of hypoglycemia, hyperglycemia, and glucose variability with morbidity and death in the pediatric intensive care unit. *Pediatrics* 2006;118(1):173–9.
- [79] Nayak PP, Davies P, Narendran P, Laker S, Gao F, Gough SC, et al. Early change in blood glucose concentration is an indicator of mortality in critically ill children. *Intensive Care Med* 2013;39(1):123–8.
- [80] Ognibene KL, Vawdrey DK, Biagas KV. The association of age, illness severity, and glycaemic status in a pediatric intensive care unit. *Pediatr Crit Care Med* 2011;12(6):e386–90.
- [81] Srinivasan V, Spinella PC, Drott HR, Roth CL, Helfaer MA, Nadkarni V. Association of timing, duration, and intensity of hyperglycemia with intensive care unit mortality in critically ill children. *Pediatr Crit Care Med* 2004;5(4): 329–36.
- [82] Leite HP, de Lima LF, de Oliveira Iglesias SB, Pacheco JC, de Carvalho WB. Malnutrition may worsen the prognosis of critically ill children with hyperglycemia and hypoglycemia. *J Parenter Enteral Nutr* 2013;37(3):335–41.
- [83] Kyle UG, Coss Bu JA, Kennedy CE, Jefferson LS. Organ dysfunction is associated with hyperglycemia in critically ill children. *Intensive Care Med* 2010;36(2): 312–20.
- [84] Preissig CM, Rigby MR. A disparity between physician attitudes and practice regarding hyperglycemia in pediatric intensive care units in the United States: a survey on actual practice habits. *Crit Care* 2010;14(1):R11.
- [85] Vlasselaers D, Milants I, Desmet L, Wouters PJ, Vanhorebeek I, van den Heuvel I, et al. Intensive insulin therapy for patients in paediatric intensive care: a prospective, randomised controlled study. *Lancet* 2009;373(9663):547–56.
- [86] Jeschke MG, Kulp GA, Kraft R, Finnerty CC, Mlcak R, Lee JO, et al. Intensive insulin therapy in severely burned pediatric patients: a prospective randomized trial. *Am J Respir Crit Care Med* 2010;182(3):351–9.
- [87] Agus MS, Steil GM, Wypij D, Costello JM, Laussen PC, Langer M, et al. Tight glycaemic control versus standard care after pediatric cardiac surgery. *N Engl J Med* 2012;367(13):1208–19.
- [88] Macrae D, Grieve R, Allen E, Sadique Z, Morris K, Pappachan J, et al. A randomized trial of hyperglycaemic control in pediatric intensive care. *N Engl J Med* 2014;370(2):107–18.
- [89] Srinivasan V, Agus MS. Tight glucose control in critically ill children—a systematic review and meta-analysis. *Pediatr Diabetes* 2014;15(2):75–83.
- [90] Faustino EV, Bogue CW. Relationship between hypoglycemia and mortality in critically ill children. *Pediatr Crit Care Med* 2010;11(6):690–8.
- [91] Hirschberg E, Larsen G, Van Duker H. Alterations in glucose homeostasis in the pediatric intensive care unit: hyperglycemia and glucose variability are associated with increased mortality and morbidity. *Pediatr Crit Care Med* 2008;9(4):361–6.
- [92] Mesotten D, Gielen M, Sterken C, Claessens K, Hermans G, Vlasselaers D, et al. Neurocognitive development of children 4 years after critical illness and treatment with tight glucose control: a randomized controlled trial. *J Am Med Assoc* 2012;308(16):1641–50.
- [93] Boluyt N, van Kempen A, Offringa M. Neurodevelopment after neonatal hypoglycemia: a systematic review and design of an optimal future study. *Pediatrics* 2006;117(6):2231–43.
- [94] Lucas A, Morley R, Cole TJ. Adverse neurodevelopmental outcome of moderate neonatal hypoglycaemia. *BMJ* 1988;297(6659):1304–8.
- [95] Tin W, Brunskill G, Kelly T, Fritz S. 15-year follow-up of recurrent “hypoglycemia” in preterm infants. *Pediatrics* 2012;130(6):e1497–503.
- [96] McKinlay CJ, Alsweiler JM, Ansell JM, Anstice NS, Chase JG, Gamble GD, et al. Neonatal glycaemia and neurodevelopmental outcomes at 2 years. *N Engl J Med* 2015;373:1507–18.
- [97] Tin W. Defining neonatal hypoglycaemia: a continuing debate. *Semin Fetal Neonatal Med* 2014;19(1):27–32.
- [98] Zhou W, Yu J, Wu Y, Zhang H. Hypoglycemia incidence and risk factors assessment in hospitalized neonates. *J Matern Fetal Neonatal Med* 2015;28(4):422–5.